



NEED HELP?



User Manual

Disclaimer: Products are intended for research use only

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SHENTEK

Residual HEK293 DNA Size Analysis Kit User Guide

Version: A/1

For Research Use Only

Product No.: 1103176

Reagents for 4×100 Reactions

Biofargo, Inc.

(IMPORTANT: Please read this document carefully before experiment.)

1. Product information

■ Product description

SHENTEK® Residual HEK293 DNA Size Analysis Kit is used to quantitate HEK293 DNA residues of different fragment sizes for various stages of biopharmaceutical products, from in-process samples to final products.

This kit utilizes real-time PCR technology to perform rapid and specific quantitation of residual HEK293 DNA fragments in samples. It is designed to amplify four different fragments (75bp, 122bp, 244bp, 562bp) for the accurate determination of their size distribution at the femtogram (fg) level. For extraction information, please refer to the SHENTEK® Residual Host Cell DNA Sample Preparation Kit User Guide (Product No. 1104191).

■ Kit contents and storage

WARNING: Please read the Material Safety Data Sheets (MSDSs) and follow the handling instructions. Wear appropriate protective eyewear, mask, clothing and gloves.

Table 1. Kit components and storage

| Reagent | Part No. | Quantity | Storage |
|-----------------------------|----------|------------------------------|------------------------------|
| qPCR Reaction Buffer | NNB001 | 850 μ L \times 8 tubes | -20°C, protect from light |
| HEK293 Primer&Probe MIX-75 | NNC012 | 300 μ L \times 1 tube | |
| HEK293 Primer&Probe MIX-122 | NNC013 | 300 μ L \times 1 tube | |
| HEK293 Primer&Probe MIX-244 | NNC014 | 300 μ L \times 1 tube | |
| HEK293 Primer&Probe MIX-562 | NNC015 | 300 μ L \times 1 tube | |
| IPC MIX | NNC069 | 550 μ L \times 1 tube | |
| DNA Dilution Buffer (DDB) | NND001 | 1.5 mL \times 3 tubes | -20°C |
| HEK293 DNA Control | NNA005 | 50 μ L \times 1 tube | |

The kit components can be stored at appropriate conditions for up to 24 months. Please check the expiration date on the labels.

■ Applied instruments, including but not limited to the following

- SHENTEK-96S Real-Time PCR System
- 7500 Real-Time PCR System
- CFX96 Real-Time PCR System
- FQD-96A Real-Time PCR System

■ Required materials not included in the kit

- Nonstick, DNase-free & Low Retention Microfuge Tubes, 1.5 mL
- Nonstick, Low Retention Tips, 1000 μ L, 100 μ L and 10 μ L
- 96-well qPCR plates or PCR 8-strip tubes

■ Related equipment

- Real-Time PCR system
- Vortex mixer
- Microplate shaker
- Micropipettes: 1000 μ L, 100 μ L and 10 μ L

■ Workflow

Serial dilutions of the control DNA preparation



Sample preparation



qPCR reaction mix preparation



qPCR amplification



Results analysis

2. Methods

■ Experiment preparation

1. Wear appropriate protective eyewear, mask, clothing and gloves.
2. Irradiate the tabletop, pipettes and tubes with UV for 30 minutes, and disinfect with 75% ethanol.
3. Thaw the kit completely at 2-8°C or melt on ice, vortex and spin briefly.

■ DNA Control serial dilutions for the standard curve

Note: The kit contains four HEK293 primer & probe mixes for different fragment lengths. Please set up four separate standard curves corresponding to each specific fragment length.

Please check the concentration on the label of the tube containing the HEK293 DNA Control prior to dilution.

Prepare a series of HEK293 DNA Control solution with DNA Dilution Buffer (DDB) and follow the serial dilution procedure below:

1. Thaw HEK293 DNA Control and DNA Dilution Buffer completely at 2-8°C or melt on ice. Vortex to mix well and quickly spin down the reagents for 3-5 seconds in microcentrifuge, and repeat 3 times.
2. Label six nonstick 1.5 mL microfuge tubes: ST0, ST1, ST2, ST3, ST4, ST5 respectively.
3. Dilute the DNA Control to 3000 pg/μL with DDB in the ST0 tube. Vortex to mix well and quickly spin down the reagents for 3-5 seconds in microcentrifuge, and repeat 3 times to mix it thoroughly.
4. Add 180 μL DDB to each tube: ST1, ST2, ST3, ST4, ST5.
5. Perform the serial dilutions:

Table 2. Dilution for HEK293 DNA Control

| Serial dilution tube | Dilution | Conc. (pg/ μ L) |
|----------------------|----------------------------------|---------------------|
| ST0 | Dilute the DNA Control with DDB | 3000 |
| ST1 | 20 μ L ST0 + 180 μ L DDB | 300 |
| ST2 | 20 μ L ST1 + 180 μ L DDB | 30 |
| ST3 | 20 μ L ST2 + 180 μ L DDB | 3 |
| ST4 | 20 μ L ST3 + 180 μ L DDB | 0.3 |
| ST5 | 20 μ L ST4 + 180 μ L DDB | 0.03 |

- *The remaining unused DDB need to be stored at 2-8°C. If the solution is cloudy or contains precipitates, heat at 37°C until it clears.*
- *At least five concentration of standard curve should be included. To select appropriate sample dilutions, we recommend to perform method validation before sample testing.*

■ sample preparation

Add 100 μ L DDB into a 1.5 mL clean centrifuge tube and label it as NCS.

NCS and samples are prepared in same way.

■ qPCR MIX preparation

1. Determine the number of reaction wells based on the standard curve, with the number of test samples and control samples. Generally, triplicates are tested for each sample.

$$\text{Number of reaction wells} = (5 \text{ standard points on the standard curve} + 1 \text{ NTC} + 1 \text{ NCS} + \text{test samples}) \times 3$$

2. Prepare qPCR MIX according to the number of reaction wells:

Table 3. qPCR MIX-75 Preparation

| Reagents | Volume/reaction | Volume for 30 reaction (includes 10% overage) |
|----------------------------|-----------------|--|
| qPCR Reaction Buffer | 15.9 μ L | 524.7 μ L |
| HEK293 Primer&Probe MIX-75 | 2.8 μ L | 92.4 μ L |
| IPC MIX | 1.3 μ L | 42.9 μ L |
| Total volume | 20 μ L | 660 μ L |

Table 4. qPCR MIX-122 Preparation

| Reagents | Volume/reaction | Volume for 30 reaction (includes 10% overage) |
|-----------------------------|-----------------|--|
| qPCR Reaction Buffer | 15.9 μ L | 524.7 μ L |
| HEK293 Primer&Probe MIX-122 | 2.8 μ L | 92.4 μ L |
| IPC MIX | 1.3 μ L | 42.9 μ L |
| Total volume | 20 μ L | 660 μ L |

Table 5. qPCR MIX-244 Preparation

| Reagents | Volume/reaction | Volume for 30 reaction (includes 10% overage) |
|-----------------------------|-----------------|--|
| qPCR Reaction Buffer | 15.9 μ L | 524.7 μ L |
| HEK293 Primer&Probe MIX-244 | 2.8 μ L | 92.4 μ L |
| IPC MIX | 1.3 μ L | 42.9 μ L |
| Total volume | 20 μ L | 660 μ L |

Table 6. qPCR MIX-562 Preparation

| Reagents | Volume/reaction | Volume for 30 reaction (includes 10% overage) |
|-----------------------------|-----------------|--|
| qPCR Reaction Buffer | 15.9 μ L | 524.7 μ L |
| HEK293 Primer&Probe MIX-562 | 2.8 μ L | 92.4 μ L |
| IPC MIX | 1.3 μ L | 42.9 μ L |
| Total volume | 20 μ L | 660 μ L |

For simultaneous detection of the four fragments, please prepare at least 120 μ L DNA template for four assays.

■ qPCR Reaction MIX preparation

1. Prepare qPCR Reaction MIX according to Table 7-10 and 96-well plates layout as shown in Table 11.

Table 7. qPCR Reaction MIX-75 Preparation

| | |
|-------------|--|
| ST-75 | 20 μ L qPCR MIX-75 + 10 μ L ST1/ST2/ST3/ST4/ ST5 |
| NTC | 20 μ L qPCR MIX-75 + 10 μ L DDB |
| NCS | 20 μ L qPCR MIX-75 + 10 μ L purified NCS |
| Test sample | 20 μ L qPCR MIX-75 + 10 μ L purified test sample |

Table 8. qPCR Reaction MIX-122 Preparation

| | |
|-------------|---|
| ST-122 | 20 μ L qPCR MIX-122 + 10 μ L ST1/ST2/ST3/ST4/ ST5 |
| NTC | 20 μ L qPCR MIX-122 + 10 μ L DDB |
| NCS | 20 μ L qPCR MIX-122 + 10 μ L purified NCS |
| Test sample | 20 μ L qPCR MIX-122 + 10 μ L purified test sample |

Table 9. qPCR Reaction MIX-244 Preparation

| | |
|-------------|---|
| ST-244 | 20 μ L qPCR MIX-244 + 10 μ L ST1/ST2/ST3/ST4/ ST5 |
| NTC | 20 μ L qPCR MIX-244 + 10 μ L DDB |
| NCS | 20 μ L qPCR MIX-244 + 10 μ L purified NCS |
| Test sample | 20 μ L qPCR MIX-244 + 10 μ L purified test sample |

Table 10. qPCR Reaction MIX-562 Preparation

| | |
|-------------|---|
| ST-562 | 20 μ L qPCR MIX-562 + 10 μ L ST1/ST2/ST3/ST4/ ST5 |
| NTC | 20 μ L qPCR MIX-562 + 10 μ L DDB |
| NCS | 20 μ L qPCR MIX-562 + 10 μ L purified NCS |
| Test sample | 20 μ L qPCR MIX-562 + 10 μ L purified test sample |

Table 11. Example of 96-well plate layout

| MIX-75 | | | MIX-122 | | | MIX-244 | | | MIX-562 | | | |
|--------|-----|-----|---------|-----|-----|---------|-----|-----|---------|-----|-----|---|
| NTC | NTC | NTC | NTC | NTC | NTC | NTC | NTC | NTC | NTC | NTC | NTC | A |
| NCS | NCS | NCS | NCS | NCS | NCS | NCS | NCS | NCS | NCS | NCS | NCS | B |
| S | S | S | S | S | S | S | S | S | S | S | S | C |
| ST5 | ST5 | ST5 | ST5 | ST5 | ST5 | ST5 | ST5 | ST5 | ST5 | ST5 | ST5 | D |
| ST4 | ST4 | ST4 | ST4 | ST4 | ST4 | ST4 | ST4 | ST4 | ST4 | ST4 | ST4 | E |
| ST3 | ST3 | ST3 | ST3 | ST3 | ST3 | ST3 | ST3 | ST3 | ST3 | ST3 | ST3 | F |
| ST2 | ST2 | ST2 | ST2 | ST2 | ST2 | ST2 | ST2 | ST2 | ST2 | ST2 | ST2 | G |
| ST1 | ST1 | ST1 | ST1 | ST1 | ST1 | ST1 | ST1 | ST1 | ST1 | ST1 | ST1 | H |
| 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 | |

- This example represents four assays, each for standard curve points of HEK293 DNA Control (ST1-ST5), 1 NTC, 1 NCS, 1 test sample and 3 replicates for each sample.
- In specific testing, the plate layout for sample loading can be adjusted based on the sample quantity. Please refer to the example shown in Table 11.

2. Seal the 96-well plate with sealing film. Mix it well in microplate shaker, then spin down the reagents for 10 seconds in centrifuge and place it in the qPCR instrument.

■ qPCR program setting

Note: The following instructions apply only to the 7500 instrument (passive reference dye ROX included) with SDS v1.4. If you use a different instrument or software, refer to the applicable instrument or software documentation.

1. Create a new document, then in the Assay drop-down list, select **Standard Curve (Absolute Quantitation)**.
2. Click **New Detector**, then enter HEK293-75 in the Name field, select **FAM** in the Reporter Dye drop-down list and select **(none)** in the Quencher Dye drop-down list, then click **OK**.
3. Create new detector for HEK293-122, HEK293-244 and HEK293-562, separately as step 2.

4. Click **New Detector**, then enter IPC in the Name field. Select **VIC** in the Reporter Dye drop-down list and select **(none)** in the Quencher Dye drop-down list, then click **OK**.
5. Select **ROX** as the passive reference dye, then Click **Next**.
6. Select the applicable set of wells for the samples, then select the corresponding detector for each well.
7. Select Finish, and then set thermal-cycling conditions:
 - a. Set the thermal cycling reaction volume to 30 µL.
 - b. Set the temperature and the time as following:

Table 12. qPCR running temperature and time

| Step | Temp. | Time(mm:sec) | Cycles |
|--------------|-------|--------------|--------|
| Activation | 95°C | 10:00 | 1 |
| Denaturation | 95°C | 00:15 | 40 |
| Annealing | 60°C | 00:30 | |
| Extension | 72°C* | 01:30 | |

*Instrument will read the fluorescence signal during this step.


8. Save the document, then click **Start** to start the real-time qPCR run.

■ Results analysis

1. Select **Set up** tab, then set tasks for each sample type by clicking on the Task Column drop-down list:
 - a. NTC: target DNA detector task = NTC
 - b. NCS, test samples: target DNA detector task = Unknown
2. Set up the standard curve as shown in the following table:

Table 13. Settings for Standard curve

| Tube label | Task | Quantity (pg/µL) |
|------------|----------|------------------|
| ST1 | Standard | 300 |
| ST2 | Standard | 30 |
| ST3 | Standard | 3 |
| ST4 | Standard | 0.3 |
| ST5 | Standard | 0.03 |

3. Select the **Results** tab, then select Amplification Plot.
4. In the Data drop-down list, select **Delta Rn vs Cycle**.
5. In the Analysis Settings window, enter the following settings :
 - a. Select **Manual Ct**.
 - b. In the Threshold field, enter 0.02.
 - c. Select **Automatic Baseline**.
6. Click the button  in the toolbar, then wait the plate analyzing.
7. Select the **Result** tab> >**Standard curve** tab, then verify the Slope, Intercept and R² values .
8. Select the Report tab, then achieve the mean quantity and standard deviation for each sample.
9. Select **File > > Export > > Results**. In the Save as type drop-down list, select **Results Export Files**, then click **Save**.
10. Set the DNA size of 75 to be 100%, calculate the percentage of the DNA size of 122, 244 and 562.
11. Analyze the Ct value of IPC. Normally, the Ct-IPC value of the sample should be within ± 1.0 of the NCS Ct-IPC value. If the Ct-IPC value of the sample is significantly higher than the Ct-IPC value of the NCS, it indicates that the sample may be inhibited. It is recommended testing the ERC samples at the same time, and take the sample recovery rate result as the criterion.
12. The Ct value of NTC and NCS should be larger than the mean Ct value of the lowest concentration in the standard curve.

Note: The parameter settings of the result analysis should be based on the specific model and the software version, and generally can also be automatically interpreted by the instrument.

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Support & Contact

The logo for SHENTEK, with the word in a bold, sans-serif font. The letters 'S', 'H', 'E', 'N', 'T', and 'E' are blue, while 'K' is green.

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